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OXOID QUALITY ASSURANCE PRODUCT SPECIFICATION

BACILLUS CEREUS AGAR BASE CM0617

BACILLUS CEREUS AGAR BASE		CM0617	
Typical Formula*			
Peptone	grams per litre	1.0	
Mannitol		10.0	
Sodium chloride		2.0	
Magnesium sulphate		0.1	
Di-sodium hydrogen phosphate		2.5	
Potassium dihydrogen phosphate		0.25	
Sodium pyruvate		10.0	
Bromothymol blue		0.12	
Agar		15.0	

^{*} adjusted as required to meet performance standards

Directions

Suspend 20.5g in 475ml of distilled water. Bring to the boil to dissolve completely. Sterilize by autoclaving at 121°C for 15 minutes. Cool to 50°C and aseptically add 25ml of Egg Yolk Emulsion (SR0047) and the contents of 1 vial of Polymyxin B Supplement (SR0099E) reconstituted as directed. Mix well and pour into sterile Petri dishes.

Physical Characteristics

Dark straw/yellow, free-flowing powder
Colour on reconstitution - dark blue
Moisture level - less than or equal to 7%
pH 7.2 ± 0.2 at 25°C
Clarity - opaque
Gel strength - firm, comparable to 15.0g/litre of agar

Microbiological Tests Using Optimum Inoculum Dilution

Control Medium: Tryptone Soya Agar

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Reactions after incubation at 37 ± 2°C for 18 ± 2 hours

Tested with the addition of 5% v/v Egg Yolk Emulsion SR0047 and Polymyxin B Supplement SR0099

Medium is challenged with 10-100 colony-forming units

Bacillus cereus ATCC®10876 3-5mm peacock blue colonies with halo Bacillus cereus ATCC®14579 3-5mm peacock blue colonies with halo

A satisfactory result is represented by recovery of positive strains equal to or greater than 70% of the control medium.

Medium is challenged with 10-100 colony-forming units

Staphylococcus aureus ATCC®25923 0.5-1mm yellow colonies, with or without halo

Enterococcus faecalis ATCC®19433 0.5-1mm yellow colonies, no halo

For Staphylococcus aureus ATCC®25923 and Enterococcus faecalis ATCC®19433, a satisfactory result is represented by recovery of equal to or greater than 50% of the control medium.

Medium is challenged with 1E+04 to 1E+06 colony-forming units

Bacillus coagulansATCC®7050No growthPseudomonas aeruginosaATCC®27853No growthSalmonella typhimuriumATCC®14028No growth

Negative strains are inhibited.

Testing performed in accordance with ISO11133:2014

Reactions after incubation at 37 ± 2°C for 21 ± 3 hours

Inoculation using diminishing sweep technique

Medium is challenged with 1E+03 to 1E+04 colony-forming units

Bacillus cereus ATCC®11778 WDCM00001 3-5mm peacock blue colonies with halo

A satisfactory result is represented by growth of positive strains with a positive diagnostic reaction (peacock blue colonies and halo).

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Reactions after incubation at 37 ± 2°C for 44 ± 4 hours

Inoculation using diminishing sweep technique

Medium is challenged with greater than 1E+03 colony-forming units

Bacillus subtilis ATCC®6633 WDCM00003 No growth or 0.5-3mm straw colonies,

no halo

A satisfactory result is represented by inhibition or growth of colonies with a negative diagnostic reaction (i.e. straw colonies without halo).

Medium is challenged with 1E+04 to 1E+06 colony-forming units

Escherichia coli ATCC®8739 WDCM00012 No growth Escherichia coli ATCC®25922 WDCM00013 No growth

Negative strains are inhibited.



Document Owner Department: QC

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Revision History

Section / Step	Description of Change	Reason for Change	Reference
Creation of ISO11133 section	Update to include testing of ISO11133:2014	Change control	BT-CC-1116
Change of S. aureus ATCC® 25923	Change <i>S. aureus</i> ATCC® 25923 on CM617 with SR47 egg yolk specification to 'with or without halos'	QC specification for CM617 is not aligned to the website information and to SR47 specification. <i>S. aureus</i> is Lecthinase +ve and can produce halos.	BT-CC-2070
Entire document	Update to new document format and correction of typographical/minor errors. Removal of 'Selective' from the document title. Change 'zone' to 'halo'.	Change Control	BT-CC-2263